

Thiol/disulfide homeostasis in patients with ankylosing spondylitis

Atalay Dogru^{1*}, Ayse Balkarli², Gozde Yildirim Cetin³, Salim Neselioglu⁴, Ozcan Erel⁴, Sevket Ercan Tunc¹, Mehmet Sahin¹

¹Department of Internal Medicine, Division of Rheumatology, Faculty of Medicine, Suleyman Demirel University, Isparta, Turkey,

²Department of Internal Medicine, Division of Rheumatology, Antalya Training and Research Hospital, Antalya, Turkey, ³Department of Internal Medicine, Division of Rheumatology, Faculty of Medicine, Kahramanmaraş Sutcu Imam University, Kahramanmaraş, Turkey,

⁴Department of Clinical Biochemistry, Faculty of Medicine, Yildirim Beyazit University, Ankara, Turkey

ABSTRACT

Ankylosing spondylitis (AS) is a chronic inflammatory disease. In many inflammatory diseases, increased production of pro-inflammatory cytokines is associated with an increase in oxidative stress mediators. Thiol/disulfide homeostasis is a marker for oxidative stress. The aim of this study was to examine the dynamic thiol/disulfide homeostasis in AS. Sixty-nine patients with AS and 60 age- and sex-matched controls were included in the study. The Bath Ankylosing Spondylitis Disease Activity Index (BASDAI) and visual analogue scale (VAS) were used to determine the disease activity. Native thiol, total thiol, and disulfide levels were measured with a novel automated method recently described by Erel and Neselioglu. The aforementioned method is also optionally manual spectrophotometric assay. The total thiol levels were significantly lower in the AS group compared with the control group ($p = 0.03$). When the patients were divided into active ($n = 35$) and inactive ($n = 34$) subgroups using BASDAI scores, the native plasma thiol and total thiol levels were significantly lower in the active AS patients compared to the inactive AS patients ($p = 0.02$, $p = 0.03$ respectively). There was a negative correlation between the plasma native thiol levels and VAS, BASDAI scores. Thiol/disulfide homeostasis may be used for elucidating the effects of oxidative stress in AS. Understanding the role of thiol/disulfide homeostasis in AS might provide new therapeutic intervention strategies for patients.

KEYWORDS: Ankylosing spondylitis; disulfide; oxidative stress; thiol

DOI: <http://dx.doi.org/10.17305/bjbms.2016.1001>

Bosn J Basic Med Sci. 2016;16(3):187-192. © 2016 ABMSFBIH

INTRODUCTION

Ankylosing spondylitis (AS) is a chronic, potentially progressive inflammatory disease involving the axial skeleton and peripheral joints [1]. While the etiopathogenesis of AS is not clearly understood, genetic, environmental, and immunological factors are thought to contribute to the disease pathogenesis [2]. Recent studies have focused on the role of oxidative stress in the pathogenesis of several inflammatory diseases including AS [3-9]. In many inflammatory diseases, increased production of pro-inflammatory cytokines is associated with an increase in oxidative stress mediators [10,11]. The equilibrium between free radical production and their elimination is known as oxidative balance. Maintaining oxidative balance limits the damage caused by free radicals. On contrary, the

imbalance in favor of free radical production results in oxidative stress. The role of oxidative stress in the pathogenesis of AS has not been clearly defined. Biasi et al. demonstrated that there is a substantial degree of neutrophil migration in active AS and that circulating neutrophils exhibit increased responsiveness to oxygen radicals in these patients [12]. A limited number of studies have investigated oxidative balance in AS patients [5,6,9,13].

Thiol, an important antioxidant, plays a key role in the eradication of reactive oxygen molecules via enzymatic and non-enzymatic pathways [14,15]. Low molecular weight thiols, such as cysteine, homocysteine, glutathione, and albumin as well as protein-thiols in particular, constitute the plasma thiol pool. Thiols engage in oxidation reactions with oxidant molecules, forming disulfide bonds. Dynamic thiol/disulfide homeostasis is essential for detoxification, regulation of signaling pathways, apoptosis, and regulation of enzymatic reactions. While abnormal thiol/disulfide concentrations are associated with many inflammatory diseases [16-22], there are no established colorimetric methods to determine thiol/disulfide

*Corresponding author: Atalay Dogru, Department of Internal Medicine, Division of Rheumatology, Faculty of Medicine, Suleyman Demirel University, 32000, Isparta/Turkey, Phone: +905063698747, Fax: +902462112830, E mail: atalay_dogru@hotmail.com

homeostasis quantitatively. Previous studies have determined thiol/disulfide homeostasis using high-performance liquid chromatography, fluorescence capillary electrophoresis, and bioluminescent systems. These methods are time-consuming, complicated, and not cost-effective [23-26]. Thus far, plasma thiol levels have typically been measured by the classic Ellman method using 5,5'-dithiobis-(2-nitrobenzoic) (DTNB) acid [27]. Today, thiol and disulfide levels can be measured individually or collectively using the new, uncomplicated, practical, and fully automatic method described by Erel and Neselioglu [28]. Since the dynamic state of oxidative balance in AS has not been investigated previously, the main goal of this study was to evaluate dynamic thiol/disulfide homeostasis in patients with AS.

MATERIALS AND METHODS

Study group and design

Sixty nine patients with AS (26 females and 43 males), who met the inclusion criteria and were diagnosed according to a modified New York classification criteria, were included in the study [29]. As the control, a group of 60 healthy, age- and sex-matched subjects, with no history of tobacco use or alcohol abuse was included. The patients were treated with non-steroidal anti-inflammatory drugs and/or anti-tumor necrosis factor (anti-TNF) agents at the time of the study. The patients were divided into subgroups according to the Bath Ankylosing Spondylitis Disease Activity Index (BASDAI), as active AS (BASDAI \geq 4) or inactive AS (BASDAI $<$ 4). The additional criteria for inclusion in the study were as follows:

- No concomitant disease (diabetes mellitus, hypertension, obesity, hyperlipidemia, depression, autoimmune disease, etc.)
- No tobacco or alcohol use
- No steroid and/or immunosuppressive treatment during the 4 weeks before blood sampling
- No infection at the time of blood sampling
- No regular use of vitamin supplements
- Completion of a written consent document.

After obtaining consent, 3 cc of venous blood was drawn from each participant into tubes containing ethylenediamine tetraacetic acid (EDTA). The samples were centrifuged, and the plasma was separated. Plasma samples were stored at -80°C before a biochemical analysis. Blood thiol disulfide homeostasis was analyzed using the new automatic method, developed by Erel and Neselioglu [28]. Disease-relevant parameters and demographic data for each study participant were recorded on a standard form designed by the researchers. The local ethics committee reviewed and approved the research protocol with a human subject.

Biochemical analysis

In this study, the modified Ellman reagent was used to quantify the total thiol content of the sample. Thus far, plasma thiol levels have typically been measured by the classic Ellman method. In a study by Erel *et al.*, the modified Ellman reagent was used to measure plasma thiol level instead of the classical Ellman reagent, and a significant correlation between the modified and original methods was found. In this study, we also demonstrated that the new method can be used instead of the classical Ellman reagent [28].

The essential principle of the Erel and Neselioglu method is the reduction of disulfide bonds (S=S) to reactive thiol groups in the presence of NaBH_4 . The reaction of formaldehyde neutralises unreactive NaBH_4 and prevents further reduction of DTNB as well as any disulfide bonds resulting from the reaction with DTNB. The modified Ellman reagent was used to quantify the total thiol content in the samples. The value of disulfide bonds is half of the difference of the native thiol content and total thiol content. The ratio of disulfide to thiol groups, the proportion of disulfide groups out of the total pool of disulfide and thiol groups, and the proportion of thiol groups relative to the total pool of disulfide and thiol groups were calculated simultaneously using the results of the reaction.

Additional measurements

The BASDAI was used to evaluate clinical disease activity, while the Bath Ankylosing Spondylitis Functional Index (BASFI) was used to determine disease-related functional insufficiency [30,31]. The visual analog scale (VAS) was used to evaluate pain [32], and spinal mobility was evaluated according to the Bath Ankylosing Spondylitis Metrology Index (BASMI) [33].

Statistical analysis

Descriptive statistics like frequency, percentage, mean, and standard deviation were used to present quantitative descriptions. The Pearson Chi-square test was used for the analysis of the association between categorical variables. For normality testing, the Shapiro-Wilks test was used for group sample sizes \leq 50, and the Kolmogorov-Smirnov test was used for group sample sizes \geq 50. The analysis of differences in continuous variables between the two groups was performed using the Mann-Whitney U-test, in cases where the data distribution was non-normal, and Student's t-test in cases where the data distribution resembled normal distribution. The associations between continuous variables were analyzed using the non-parametric Spearman correlation test in cases where the data distribution was not consistent with normal distribution.

The Pearson correlation test was used for continuous variables that showed a normal distribution. All statistical analyses were conducted using Statistical Package for the Social Sciences 13.0 (SPSS 13.0). Probability value of $p < 0.05$ was considered statistically significant.

RESULTS

The mean age of the AS patients (43 males, 62.3%) was 39 ± 10.58 years, and the mean age of the control group (37 males, 61.7%) was 38 ± 10.56 years. The patient and control groups were similar given the age and sex ($p > 0.05$) (Table 1). The patient follow-up time was 4 (1.2-7.5) years, and the median duration of symptoms was 6 (2.6-11.2) years. The laboratory and disease-related data of the AS patients are listed in Table 2.

While there were no differences in native thiol levels between the AS patients and the control group (327.2 ± 52.118 vs. 338.9 ± 39.23 $\mu\text{mol/L}$, $p = 0.081$), total thiol levels were significantly lower in the AS patients group compared

to the control group (363.5 ± 56.99 vs. 375.95 ± 41.92 $\mu\text{mol/L}$, $p = 0.03$) (Table 3). In addition, there were significant differences between the AS patients and the control group regarding plasma disulfide levels (14.55 ± 7.2 vs. 17.07 ± 7.11 $\mu\text{mol/L}$, $p = 0.01$). Ratios obtained using plasma native thiol, total thiol, and disulfide levels differed significantly between the AS patients and the control group. While the disulfide/native thiol and disulfide/total thiol ratios were significantly higher in the control group, the native thiol/total thiol ratio was higher in the AS patients ($p = 0.03$, 0.03 , and 0.03 , respectively) (Table 4). When the AS patients group was evaluated by active ($n = 35$) and inactive ($n = 34$) AS subgroups according to the BASDAI score, the plasma native thiol (299.7 ± 45.12 vs. 338.25 ± 54.23 $\mu\text{mol/L}$, $p = 0.02$) and total thiol (330.8 ± 50.11 vs. 369.2 ± 59.32 $\mu\text{mol/L}$, $p = 0.03$) levels were lower among the active AS patients compared to the inactive AS patients (Table 4).

There was a statistically significant negative relationship between the native and total thiol concentrations and VAS ($\rho = -0.351$; $p = 0.004$ and $\rho = -0.369$; $p = 0.002$). A statistically significant negative correlation between the native and total thiol and BASDAI ($r = -0.388$, $p = 0.001$ and $r = -0.383$, $p = 0.001$) was also identified. A negative statistically significant correlation was established between the age and native and total levels of thiol. No significant relationship was established between the thiol/disulfide values and erythrocyte sedimentation rate (ESR), C-reactive protein (CRP), BASMI, and BASFI (Table 5).

When the AS patients were classified into subgroups according to whether they received anti-TNF treatment

TABLE 1. Baseline characteristics of study groups

Baseline characteristics	AS (n=69)	Control (n=60)	p value
Age (SD) (years)	39 (10)	38 (10)	NS
Male, n (%)	43 (62.3)	37 (61.7)	NS

Values are presented as mean (SD). NS: non-significant; AS: Ankylosing spondylitis; SD: Standard deviation

TABLE 2. Demographic and laboratory data of AS patients

Parameter	Result
Fasting blood glucose (74-106 mg/dl)	87 (7.5)**
Urea (20-50 mg/dl)	26 (8)**
Creatinine (0.66-1.09 mg/dl)	0.8 (0.1)**
AST (0-31 U/L)	20 (6)**
ALT (0-34 U/L)	19 (7)**
ESR (mm/h)	18 (10-37)*
C-reactive protein (0-3 mg/L)	6 (3.4-13.1)*
WBC (5200-12,400 $10^3/\mu\text{L}$)	7.427 (1.897)**
Hemoglobin (13.6-17.2 g/dl)	13 (1.9)**
Platelets (156-373 $10^3/\mu\text{L}$)	275.666 (58.3)**
Onset of first symptoms, years	6.0 (2.6-11.2)*
Duration of disease, years	4 (1.2-7.5)*
Uveitis, n (%)	11 (15.9)**
Peripheral arthritis, n (%)	25 (36.2)**
HLA B27, n (%)	54 (78.3)**
BASDAI	4 (2.5-5.0)*
VAS	4.51 (1.95)**
BASMI	1 (0-4)*
BASFI	3.2 (0.7-5.1)*

*Values are presented as median (25-75 interquartile ranges), ** Values are presented as mean (standard deviation), AST: Aspartate aminotransferase; ALT: Alanine transaminase; ESR: Erythrocyte sedimentation rate; WBC: White blood cell; HLA: Human leukocyte antigen; BASDAI: Bath Ankylosing Spondylitis Disease Activity Index; VAS: Visual Analog Scale; BASMI: Bath Ankylosing Spondylitis Metrology Index; BASFI: Bath Ankylosing Spondylitis Functional Index

TABLE 3. Plasma thiol-disulfide levels of AS and control group

Parameter	AS (n=69)	Control (n=60)	p value
Native thiol, $\mu\text{mol/L}$	327 (52)	338 (39)	NS
Total thiol, $\mu\text{mol/L}$	363 (56)	375 (41)	0.03*
Disulfide, $\mu\text{mol/L}$	14 (7.2)	17 (7.1)	0.01*
Disulfide/native thiol, %	4.2 (2.3)	5.1 (2.2)	0.03*
Disulfide/total thiol, %	3.9 (1.8)	4.6 (1.8)	0.03*
Native thiol/total thiol, %	92 (3.7)	90 (3.6)	0.03*

Values are presented as mean \pm standard deviation.

* $p < 0.05$. NS: Non-significant; AS: Ankylosing spondylitis

TABLE 4. Plasma thiol-disulfide levels according to disease activity score (BASDAI)

Parameter	BASDAI ≥ 4 (n=35)	BASDAI < 4 (n=34)	p value
Native thiol ($\mu\text{mol/L}$)	299 (45)	338 (54)	0.02*
Total thiol ($\mu\text{mol/L}$)	330 (50)	369 (59)	0.03*
Disulfide ($\mu\text{mol/L}$)	14 (7.2)	14 (7.2)	NS
Disulfide/native thiol (%)	4.7 (2.3)	4.2 (2.3)	NS
Disulfide/total thiol (%)	4.3 (1.9)	3.8 (1.8)	NS
Native thiol/total thiol (%)	91 (3.8)	92 (3.6)	NS

Values are presented as mean \pm standard deviation. * $p < 0.05$.

NS: Non-significant; BASDAI: Bath Ankylosing Spondylitis Disease Activity Index

TABLE 5. Correlation analysis between thiol/disulfide and other clinical or biochemical parameters

Parameter	Native thiol (rho)	Total thiol (rho)	Disulfide (rho)	Disulfide/native thiol (rho)	Disulfide/total thiol (rho)	Native thiol/total thiol (rho)
Age	-0.470**	-0.450**	-0.112	0.081	0.081	0.081
BASDAI	-0.388**	-0.383**	-0.142	-0.008	-0.008	-0.008
VAS	-0.351**	-0.369**	-0.183	-0.033	-0.033	-0.033
BASMI	-0.242	-0.202	0.34	0.89	0.89	0.89
BASFI	-0.234	-0.245	-0.060	0.027	0.027	0.027
ESR	-0.076	-0.089	-0.108	-0.075	-0.075	-0.075
C-reactive protein	-0.123	-0.188	-0.191	-0.143	-0.143	-0.143

*0.01 < p < 0.05, **0.001 < p < 0.01. ESR: Erythrocyte sedimentation rate; BASDAI: Bath Ankylosing Spondylitis Disease Activity Index; VAS: Visual analog scale; BASMI: Bath Ankylosing Spondylitis Metrology Index; BASFI: Bath Ankylosing Spondylitis Functional Index

($n = 34$), there was no significant difference in the plasma native thiol, total thiol, or disulfide concentrations between the subgroups ($p > 0.05$).

DISCUSSION

In the current study, thiol, an important component of the plasma antioxidant system, was significantly reduced in AS. Especially in the active AS patients, the thiol levels were significantly lower. In addition, there was a statistically significant negative correlation between the plasma thiol concentrations and the VAS, BASDAI scores. During the inactive period, the thiol/disulfide values of the controls were similar as the values of the patients. This result demonstrates that antioxidant systems are affected in inflammatory diseases, especially during the active period. The inflammatory response outside of the activation period does not continue. As the result, thiol can be used clinically as an activation marker. In addition, thiol can be helpful in the evaluation of patients' treatment as well as the BASDAI score. In a study by Il'in *et al.*, including 240 patients with rheumatological diseases, oxygen-dependent neutrophil metabolism and superoxide anion production were elevated in AS patients [34]. Karakoc *et al.* similarly reported a higher level of oxidative stress in AS patients compared to control subjects [9]. Yazici *et al.* reported high levels of neutrophil activation-dependent oxidative stress and low thiol concentrations in AS patients, similar to the findings in this study. Low thiol levels were observed in both active and inactive AS patients. Another study showed that antioxidant capacity in synovial fluid and thiol oxidation were diminished, and the authors proposed that this effect might be a significant component of AS pathophysiology [6]. In a study which investigated thiol levels in chronic renal disease (CRD), thiol levels were significantly reduced in CRD and hemodialysis patients compared to a control group [35]. Kundi *et al.* demonstrated that thiol levels correlated with the severity of coronary atherosclerosis and the mortality rate was higher in the group with low thiol levels [36]. Furthermore, thiol levels were shown to be markedly reduced in patients with migraine [37]. In a study carried out by Erkus *et al.*, a connection between low plasma thiol

levels and left ventricular diastolic dysfunction was reported. The authors suggested that increased thiol levels could play a protective role in diastolic dysfunction [38].

The oxidation of reactive oxygen radicals results in the formation of disulfide bonds. Disulfide bonds can revert to thiol groups, a mechanism by which thiol/disulfide homeostasis is maintained [39,40]. Abnormal thiol/disulfide balance is associated with many diseases [16-22]. In a recent study, including patients with pre-diabetes, Ates *et al.* showed a positive correlation between disulfide and blood glucose concentrations [41]. In a study, including patients with newly diagnosed primary hypertension, there was a positive correlation between disulfide levels and blood pressure. Disulfide was identified as an independent risk factor for hypertension in this study [42]. In another study, an increase was found in a disulfide/thiol ratio in patients with idiopathic recurrent pregnancy loss; however, there was no difference in the disulfide levels [43]. In our study, although elevated plasma disulfide levels in the AS patients were expected to be observed, lower plasma thiol concentrations in the AS group compared to the control group, were detected. The low values in disulfide resulted in the disulfide/native thiol and disulfide/total thiol values being low compared to the controls. The disulfide levels were similar when evaluated according to the disease activation, which might be explained by several factors. First, oxidative stress in AS might occur through a mechanism that is independent of disulfide levels. Second, the thiol/disulfide homeostasis system might be insufficient in AS patients. This condition could also result from a disruption of the oxidant/antioxidant balance in AS and the diminished antioxidant response. Additional studies with large patient populations might help clarify this issue.

CONCLUSION

The new colorimetric method, defined by Erel and Neselioglu, for measuring thiol/disulfide homeostasis was used in this study [28]. Thiol chemistry is a rapidly expanding field in basic and applied molecular life sciences. The measurement of thiol/disulfide homeostasis is critical for elucidating the effects of oxidative stress and for evaluating disease

activation. Many articles in the literature related to oxidative stress demonstrated conflicting data. Different authors have tried to establish a connection between oxidative stress and pathophysiology of disease. However, in different pathophysiological conditions, oxidant molecules are found to be high and antioxidant molecules are found to be low in general. This leads to the information pollution and impedes access to accurate information about this subject. Randomized controlled trials with large patient populations are necessary to continue the research in this field.

DECLARATION OF INTERESTS

The authors declare no conflict of interests.

REFERENCES

- [1] Khan MA. Ankylosing spondylitis: Clinical features of ankylosing spondylitis. In: Hochberg MC, Silman AJ, Smolen JS, Weinblat ME, Weisman MH, editors. *Rheumatology*, 3rd ed. Spain: Mosby; 2003. p. 1161-81.
- [2] Khan MA, Ball EJ. Genetic aspects of ankylosing spondylitis. *Best Pract Res Clin Rheumatol* 2002;16(4):675-90. <http://dx.doi.org/10.1053/berh.2002.0243>. [http://dx.doi.org/10.1016/S1521-6942\(02\)90243-3](http://dx.doi.org/10.1016/S1521-6942(02)90243-3)
- [3] Köse K, Yazici C, Cambay N, Ascioğlu O, Dogan P. Lipid peroxidation and erythrocyte antioxidant enzymes in patients with Behçet's disease. *Tohoku J Exp Med* 2002;197(1):9-16. <http://dx.doi.org/10.1620/tjem.197.9>.
- [4] Karatas F, Ozates I, Canatan H, Halifeoglu I, Karatepe M. Colakt R. Antioxidant status and lipid peroxidation in patients with rheumatoid arthritis. *Indian J Med Res* 2003;118:178-81.
- [5] Ozgocmen S, Sogut S, Ardicoglu O, Fadillioglu E, Pekcutucu I, Akyol O. Serum nitric oxide, catalase, superoxide dismutase, and malondialdehyde status in patients with ankylosing spondylitis. *Rheumatol Int* 2004;24(2):80-3. <http://dx.doi.org/10.1007/s00296-003-0335-y>.
- [6] Yazici C, Köse K, Calis M, Kuzugüden S, Kirnap M. Protein oxidation status in patients with ankylosing spondylitis. *Rheumatology (Oxford)* 2004;43(10):1235-9. <http://dx.doi.org/10.1093/rheumatology/keh317>.
- [7] Yıldırım K, Karatay S, Güreşer G, Kızıltunç A, Uğur M, Şenel K. Antioxidant enzymes capacity in patients with rheumatoid arthritis: The relationship with disease activity score. *Romatizma* 2004;19:81-6.
- [8] Sarban S, Kocyigit A, Yazar M, Isikan UE. Plasma total antioxidant capacity, lipid peroxidation, and erythrocyte antioxidant enzyme activities in patients with rheumatoid arthritis and osteoarthritis. *Clin Biochem* 2005;38(11):981-6. <http://dx.doi.org/10.1016/j.clinbiochem.2005.08.003>.
- [9] Karakoc M, Altındag O, Keles H, Soran N, Selek S. Serum oxidative-antioxidative status in patients with ankylosing spondylitis. *Rheumatol Int* 2007;27(12):1131-4. <http://dx.doi.org/10.1007/s00296-007-0352-3>.
- [10] LeGrand A, Fermor B, Fink C, Pisetsky DS, Weinberg JB, Vail TP, et al. Interleukin-1, tumor necrosis factor alpha, and interleukin-17 synergistically up-regulate nitric oxide and prostaglandin E2 production in explants of human osteoarthritic knee menisci. *Arthritis Rheum* 2001;44(9):2078-83. [http://dx.doi.org/10.1002/1529-0131\(200109\)44:9<2078:AID-ART358>3.0.CO;2-J](http://dx.doi.org/10.1002/1529-0131(200109)44:9<2078:AID-ART358>3.0.CO;2-J).
- [11] Horton JW. Free radicals and lipid peroxidation mediated injury in burn trauma: The role of antioxidant therapy. *Toxicology* 2003;189(1-2):75-88. [http://dx.doi.org/10.1016/S0300-483X\(03\)00154-9](http://dx.doi.org/10.1016/S0300-483X(03)00154-9).
- [12] Biasi D, Carletto A, Caramaschi P, Bellavite P, Andrioli G, Caraffi M, et al. Neutrophil functions, spondylarthropathies and HLA-B27: A study of 43 patients. *Clin Exp Rheumatol* 1995;13(5):623-7.
- [13] Stichtenoth DO, Wollenhaupt J, Andersone D, Zeidler H, Frölich JC. Elevated serum nitrate concentrations in active spondylarthropathies. *Br J Rheumatol* 1995;34(7):616-9. <http://dx.doi.org/10.1093/rheumatology/34.7.616>.
- [14] Cadenas E. Biochemistry of oxygen toxicity. *Annu Rev Biochem* 1989;58:79-110. <http://dx.doi.org/10.1146/annurev.bi.58.070189.000455>.
- [15] Young IS, Woodside JV. Antioxidants in health and disease. *J Clin Pathol* 2001;54(3):176-86. <http://dx.doi.org/10.1136/jcp.54.3.176>.
- [16] Biswas S, Chida AS, Rahman I. Redox modifications of protein-thiols: Emerging roles in cell signaling. *Biochem Pharmacol* 2006;71(5):551-64. <http://dx.doi.org/10.1016/j.bcp.2005.10.044>.
- [17] Circu ML, Aw TY. Reactive oxygen species, cellular redox systems, and apoptosis. *Free Radic Biol Med* 2010;48(6):749-62. <http://dx.doi.org/10.1016/j.freeradbiomed.2009.12.022>.
- [18] Matteucci E, Giampietro O. Thiol signalling network with an eye to diabetes. *Molecules* 2010;15(12):8890-903. <http://dx.doi.org/10.3390/molecules15128890>.
- [19] Tetik S, Ahmad S, Alturfan AA, Fresko I, Disbudak M, Sahin Y, et al. Determination of oxidant stress in plasma of rheumatoid arthritis and primary osteoarthritis patients. *Indian J Biochem Biophys* 2010;47(6):353-8.
- [20] Go YM, Jones DP. Cysteine/cystine redox signaling in cardiovascular disease. *Free Radic Biol Med* 2011;50(4):495-509. <http://dx.doi.org/10.1016/j.freeradbiomed.2010.11.029>.
- [21] Kuo LM, Kuo CY, Lin CY, Hung MF, Shen JJ, Hwang TL. Intracellular glutathione depletion by oridonin leads to apoptosis in hepatic stellate cells. *Molecules* 2014;19(3):3327-44. <http://dx.doi.org/10.3390/molecules19033327>.
- [22] Prabhu A, Sarcar B, Kahali S, Yuan Z, Johnson JJ, Adam KP, et al. Cysteine catabolism: A novel metabolic pathway contributing to glioblastoma growth. *Cancer Res* 2014;74(3):787-96. <http://dx.doi.org/10.1158/0008-5472.CAN-13-1423>.
- [23] Mourad T, Min KL, Steghens JP. Measurement of oxidized glutathione by enzymatic recycling coupled to bioluminescent detection. *Anal Biochem* 2000;283(2):146-52. <http://dx.doi.org/10.1006/abio.2000.4659>.
- [24] Carru C, Deiana L, Sotgia S, Pes GM, Zinellu A. Plasma thiols redox status by laser-induced fluorescence capillary electrophoresis. *Electrophoresis* 2004;25(6):882-9. <http://dx.doi.org/10.1002/elps.200305768>.
- [25] Chen W, Zhao Y, Seefeldt T, Guan X. Determination of thiols and disulfides via HPLC quantification of 5-thio-2-nitrobenzoic acid. *J Pharm Biomed Anal* 2008;48(5):1375-80. <http://dx.doi.org/10.1016/j.jpba.2008.08.033>.
- [26] Glowacki R, Bald E. Fully automated method for simultaneous determination of total cysteine, cysteinylglycine, glutathione and homocysteine in plasma by HPLC with UV absorbance detection. *J Chromatogr B Analyt Technol Biomed Life Sci* 2009;877(28):3400-4. <http://dx.doi.org/10.1016/j.jchromb.2009.06.012>.
- [27] Winther JR, Thorpe C. Quantification of thiols and disulfides. *Biochim Biophys Acta* 2014;1840(2):838-46. <http://dx.doi.org/10.1016/j.bbagen.2013.03.031>.
- [28] Erel O, Neselioglu S. A novel and automated assay for thiol/disulphide homeostasis. *Clin Biochem* 2014;47(18):326-32. <http://dx.doi.org/10.1016/j.clinbiochem.2014.09.026>.
- [29] Van der Linden S, Valkenburg HA, Cats A. Evaluation of diagnostic criteria for ankylosing spondylitis. A proposal for modification of the New York criteria. *Arthritis Rheum* 1984;27(4):361-8. <http://dx.doi.org/10.1002/art.1780270401>.
- [30] Garrett S, Jenkinson T, Kennedy LG, Whitelock H, Gaisford P, Calin A. A new approach to defining disease status in ankylosing

- spondylitis: The Bath Ankylosing Spondylitis Disease Activity Index. *J Rheumatol* 1994;21(12):2286-91.
- [31] Calin A, Garrett S, Whitelock H, Kennedy LG, O'Hea J, Mallorie P, et al. A new approach to defining functional ability in ankylosing spondylitis: The development of the bath ankylosing spondylitis functional index. *J Rheumatol* 1994;21(12):2281-5.
- [32] Price DD, McGrath PA, Rafii A, Buckingham B. The validation of visual analogue scales as ratio scale measures for chronic and experimental pain. *Pain* 1983;17(1):45-56.
[http://dx.doi.org/10.1016/0304-3959\(83\)90126-4](http://dx.doi.org/10.1016/0304-3959(83)90126-4).
- [33] Jenkinson TR, Mallorie PA, Whitelock HC, Kennedy LG, Garrett SL, Calin A. Defining spinal mobility in ankylosing spondylitis (AS). The Bath AS metrology index. *J Rheumatol* 1994;21(9):1694-8.
- [34] Il'in MV, Mal'tseva PA, Rosanov DV, Volkova AS, Khrustalev AO. Changes in oxidative stress and apoptosis parameters of neutrophils in rheumatoid diseases. *Zh Mikrobiol Epidemiol Immunobiol* 2012;(1):89-92.
- [35] Himmelfarb J, McMonagle E, McMenamin E. Plasma protein thiol oxidation and carbonyl formation in chronic renal failure. *Kidney Int* 2000;58(6):2571-8.
<http://dx.doi.org/10.1046/j.1523-1755.2000.00443.x>.
- [36] Kundi H, Erel Ö, Balun A, Çiçekçioğlu H, Cetin M, Kiziltunç E, et al. Association of thiol/disulfide ratio with syntax score in patients with NSTEMI. *Scand Cardiovasc J* 2015;49(2):95-100.
<http://dx.doi.org/10.3109/14017431.2015.1013153>.
- [37] Eren Y, Dirik E, Neselioglu S, Erel Ö. Oxidative stress and decreased thiol level in patients with migraine: Cross-sectional study. *Acta Neurol Belg* 2015;115(4):643-9.
<http://dx.doi.org/10.1007/s13760-015-0427-y>.
- [38] Erkus ME, Altıparmak IH, Akyuz AR, Demirbag R, Sezen Y, Günebakmaz O, et al. The association between plasma thiol levels and left ventricular diastolic dysfunction in patient with hypertension. *Scand J Clin Lab Invest.* 2015;75(8):667-73.
<http://dx.doi.org/10.1016/j.amjcard.2015.01.076>.
- [39] Jones DP, Liang Y. Measuring the poise of thiol/disulfide couples *in vivo*. *Free Radic Biol Med* 2009;47(10):1329-38.
<http://dx.doi.org/10.1016/j.freeradbiomed.2009.08.021>.
- [40] Cremers CM, Jakob U. Oxidant sensing by reversible disulfide bond formation. *J Biol Chem* 2013;288(37):26489-96.
<http://dx.doi.org/10.1074/jbc.R113.462929>.
- [41] Ates I, Kaplan M, Inan B, Alisik M, Erel O, Yilmaz N, et al. How does thiol/disulfide homeostasis change in prediabetic patients? *Diabetes Res Clin Pract* 2015;110(2):166-71.
<http://dx.doi.org/10.1016/j.diabres.2015.09.011>.
- [42] Ates I, Ozkayar N, Inan B, Yilmaz FM, Topcuoglu C, Neselioglu S, et al. Dynamic thiol/disulphide homeostasis in patients with newly diagnosed primary hypertension. *J Am Soc Hypertens* 2016;10(2):159-66.
<http://dx.doi.org/10.1016/j.jash.2015.12.008>.
- [43] Erkenekli K, Sanhal CY, Yucel A, Bicer CK, Erel O, Uygur D. Thiol/disulfide homeostasis in patients with idiopathic recurrent pregnancy loss assessed by a novel assay: Report of a preliminary study. *J Obstet Gynaecol Res* 2016;42(2):136-41.
<http://dx.doi.org/10.1111/jog.12860>.