# Predictors of COVID-19 severity among pregnant patients

Marcin Januszewski<sup>1</sup>, Laura Ziuzia-Januszewska<sup>2</sup>, Alicja A. Jakimiuk<sup>3</sup>, Tomasz Oleksik<sup>1</sup>, Marek Pokulniewicz<sup>1</sup>, Waldemar Wierzba<sup>1,4</sup>, Krzysztof Kozłowski<sup>5</sup>, Artur J. Jakimiuk<sup>1,6\*</sup>

# ABSTRACT

Coronavirus disease 2019 (COVID-19) was declared a pandemic and has spread around the globe, unsparingly affecting vulnerable populations. Effective prevention measures for pregnant women, who are particularly affected, include early identification of those patients at risk of developing in-hospital complications, and the continuous improvement of maternal-fetal treatment strategies to ensure the efficient use of health resources. The objective of our retrospective study was to determine which patient biomarkers on hospital admission correlate with disease severity as measured by disease course classification, the need for oxygen supplementation and higher demand for oxygen, the need for mechanical ventilation, intensive care unit admission, and length of hospital stay. Analysis of 52 PCR SARS-CoV-2 positive pregnant women revealed that the median date of hospital admission was the 30<sup>th</sup> gestational week, with dyspnea, cough, and fever as the leading symptoms. The presence of diabetes and hypertension predisposed pregnant women to the severe course of illness. Lung involvement shown by CT scans on admission correlated with the greater clinical severity. The main laboratory predictors of disease progression were lymphocytopenia, hypocalcemia, low total cholesterol, low total protein levels, and high serum levels of C-reactive protein, ferritin, interleukin-6, glucose, lactate dehydrogenase, procalcitonin, and troponin I. Further, research with a larger cohort of pregnant women is needed to determine the utility of these results for everyday practice.

KEYWORDS: COVID-19; pregnancy; SARS-CoV-2; clinical course; predictors; disease severity; lymphocytopenia; hypocalcemia; low total protein; inflammation biomarkers

## INTRODUCTION

Coronavirus disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is widespread and globally claims more victims with each passing month [1]. As more alarming data about the characteristics of new SARS-CoV-2 variants emerge and as we witness the natural evolution and increased infection rates of COVID-19, it is increasingly important to be able to prioritize critical care

<sup>4</sup>University of Humanities and Economics in Lodz, Satellite Campus in Warsaw, Warsaw, Poland

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services in situations, where the number of patients may be overwhelming. Prenatal care, which is particularly affected, deserves special attention and continuous improvement of its treatment strategies.

The distribution of disease severity in pregnant women is similar to the distribution seen in non-pregnant populations, with 86% of pregnant women manifesting mild disease, 9% severe, and 5% critical [2].

SARS-CoV-2 affects nearly every organ system [3-7], as well as affecting the mental health of both infected and non-infected pregnant women [8]. Leading symptoms include fever (88.7%), cough (67.8%), fatigue (38%), and the over-production of mucus (33.7%) [4]. Severe COVID-19 is characterized by the development of acute respiratory distress syndrome (ARDS), hypotensive shock, and multiorgan failure and requires the patient's admission to intensive care unit (ICU) and mechanical ventilation [3-5].

Severe disease risk factors include comorbidities, advanced age, male sex, obesity, and genetic predispositions [3,4,5,9]. Severe COVID-19 is mainly an immune-mediated disorder triggered by the SARS-CoV-2 infection promoting excessive inflammation and hypercoagulable states [10].

During pregnancy, physiological adaptations of the respiratory tract, immunomodulation, hypercoagulability, processes that increase insulin resistance, and the development of hypertension, predispose SARS-CoV-2-infected women

<sup>&</sup>lt;sup>1</sup>Department of Obstetrics and Gynecology, Central Clinical Hospital of the Ministry of the Interior and Administration, Warsaw, Poland, <sup>2</sup>Department of Otolaryngology, Central Clinical Hospital of the Ministry of Interior and Administration, Warsaw, Poland, <sup>3</sup>Department of Plastic Surgery, Central Clinical Hospital of the Ministry of the Interior and Administration, Warsaw, Poland,

<sup>&</sup>lt;sup>5</sup>Department of Constitutional Law, Jagiellonian University in Krakow, Krakow, Poland,

<sup>6</sup>Center for Reproductive Health, Institute of Mother and Child, Warsaw, Poland

<sup>\*</sup>Corresponding author: Artur J. Jakimiuk, Center for Reproductive Health, Institute of Mother and Child, Warsaw, Poland. E-mail: jakimiuk@yahoo.com

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toward a severe course of illness, leading to maternal and fetal mortality and morbidity [10-14].

Data emerging from meta-analyses in the literature show that pregnant women may have an increased risk of developing severe symptoms and a higher risk of pneumonia, ICU admission, the requirement for invasive ventilation and extracorporeal membrane oxygenation (ECMO), and death [15-17].

Moreover, serious adverse outcomes have been observed among pregnant women with previous coronavirus infections, namely, severe acute respiratory syndrome (SARS) and Middle East respiratory syndrome (MERS) [18], influenza [19,20], and respiratory syncytial virus [21].

There is currently no prognostic biomarker available to identify pregnant patients who are at imminent risk of a severe course of COVID-19, with all associated maternal and fetal complications, and who require immediate medical attention.

The objective of our study was to determine to determine, in which patient characteristics and laboratory results on hospital admission correlate with disease severity as measured by disease course classification, the need for oxygen supplementation and higher demand, the need for mechanical ventilation, ICU admission, and length of hospital stay.

## MATERIALS AND METHODS

#### Study population

This retrospective single-center study was undertaken in the Department of Obstetrics and Gynecology, at the Central Clinical Hospital of the Ministry of the Interior and Administration in Warsaw, Poland. The study group comprised 52 pregnant women with COVID-19 who had been admitted for treatment between 15 May 2020 and 26 April 2021.

Inclusion criteria, similar to admission indications for pregnant women with COVID-19, were temperature >39°C despite the use of acetaminophen, respiratory rate >30/min,  $SpO_2 <95\%$  measured at time of admission without oxygen supplementation, patient requiring oxygen, and critical disease. COVID-19 was confirmed using a PCR test prior to admission.

Exclusion criteria were that the patient was admitted to hospital for obstetric and/or other non-COVID-19-related reasons.

#### Clinical course of the disease

According to the guidelines of the Polish Association of Epidemiologists and Infectiologists, patients were divided into four cohorts based on the severity of their symptoms and test results, corresponding to the relative course of their illness: mild, moderate, severe, and critical [22].

Mild cases were characteristically clinically stable with mild upper respiratory tract symptoms. Moderate cases included clinical indicators as well as lung involvement shown on imaging. Patients in the severe cohort demonstrated respiratory failure and peripheral SpO<sub>2</sub> <90%. Those in the critical cohort were characterized by ARDS, hypotensive shock, multiorgan failure, and loss of consciousness [22].

#### Study procedures

On admission, all women underwent complete blood biochemistry and urine tests, a coagulation profile, and in cases where moderate, severe, or critical forms of COVID-19 were suspected, a CT chest scan (without contrast) was performed.

We analyzed the following data: age of patient, body mass index (BMI), gestational age, initial vital signs and symptoms, pre-existing comorbidities such as diabetes mellitus, hypertension, hypothyroidism, asthma, and any history of smoking.

#### Ethical statement

The research project was approved by the local Bioethics Committee (Decision Number104/2021).

#### Statistical analysis

We used Statistica 13.3 (StatSoft Poland) for our data analysis. Mean values and standard deviations were used to describe the study groups. In case of skewed distributions, the median was calculated as a measure of central tendency, and the scatter of data was shown in relation to the 25<sup>th</sup> and 75<sup>th</sup> percentiles. Qualitative variables were presented as percentages. Spearman's rank correlation was used to assess correlation. In case of qualitative variables, the Chi-square test was used to compare the frequencies of the studied characteristics. Differences were considered statistically significant at p < 0.05. Logistic regression was performed to analyze the association of patient characteristics and laboratory parameters and the risk of severe-to-critical disease. Non-linear data were categorized. Variables with more than 20% of their values missing were not considered in this analysis, in other cases, missing values were analyzed as a separate category. Comparisons between the four disease severity groups were performed using the Kruskal-Wallis test followed by pair-wise comparison using Dunn's post hoc test for continuous variables, and Pearson's chi-square test for categorical variables. As there was only one patient with a critical course of the disease, the severe and critical groups were combined in the analysis as a new grouping, severe-to-critical disease.

## RESULTS

Gestational age ranged from 17 to 37 weeks. Four patients were at 17-22 weeks of gestation, 14 were at 24-28, 17 were at 29-33, 15 were at 34-36, and 2 patients were at >37 weeks of gestation. The mean age of the patients was  $31.9 \pm 4.79$  years. Mean BMI at admission was 28.36 (9.88) kg/m<sup>2</sup>. None of the patients reported a history of smoking. Symptoms on admission were: dyspnea (n = 48, 92.31%), cough (n = 47, 90.38%), fever (n = 33, 63.46%), fatigue and muscle aches (n = 22, 42.31\%), smell and taste disorders (n = 14, 26.92%), headache (n = 12, 23.08%), sore throat (n = 6, 11.54%), and nasal discharge (n = 5, 9.62%). Coexisting diseases were diabetes (n = 9, 17.65%), hypertension (n = 5, 10.00%), hypothyroidism (n = 18, 35.29%), and asthma (n = 2, 3.85%). Mild, moderate, severe, and critical COVID-19 accounted for n = 9 (17.31%), n = 25 (48.08%), n = 17 (32.69%), and n = 1 (1.92%) cases, respectively. The main outcomes measured were length of hospitalization (median = 8 [range = 2-23] days), the need for oxygen supplementation (n = 42, 80.77%), median oxygen flow rate (median = 4 [range = 0-15]), requirement for high-flow therapy (n = 9, 17.31%), and the need for ICU admission (n = 2, 3.85%). There were no cases of tracheal intubation, mechanical ventilation, or ECMO. The median lung involvement seen by CT imaging was 20% (IQR = 11), ranging from 1% to 60%. The most common abnormalities shown in the laboratory results that were elevated C-reactive protein (CRP) (94.23%), elevated D-dimer (90.63%), elevated interleukin 6 (IL-6) (88.46%), elevated fibrinogen (88%), hypoproteinemia (66.67%), decreased vitamin D (62.22%), elevated lactate dehydrogenase (LDH) (56%), hyperglycemia (48.78%), anemia (48.08%), elevated alkaline phosphatase (ALP) (46.15%), elevated aspartate aminotransferase (AST) (40.38%), lymphopenia (38.46%), neutrophilia (30.77%), elevated alanine transaminase (ALT) (30%), and elevated bile acids (35.71%). Data regarding patients' characteristics, clinical course parameters, and laboratory abnormalities are presented in Table 1.

#### Main predictors of severe course of illness

Diabetes as a comorbidity was correlated with the need for high-flow oxygen therapy and higher oxygen flow. Hypertension was correlated with oxygen flow demand during hospitalization. The percentage of lung involvement correlated with four of the six main outcomes: the severity of the course of the COVID-19, the oxygen flow (l/min), the need for high-flow oxygen therapy, and the need for ICU admission (Table 2).

Lymphocytopenia, low levels of serum calcium, total cholesterol and total protein levels, high levels of serum

**TABLE 1.** Clinical characteristics of 52 pregnant COVID-19 patients

Variable	Cases
Age, years (n=52)	
Mean±SD	31.9±4.79
Hbd, weeks (n=52)	
Median (IQR)	30 (7)
Hbd, ranges, n (%); (n=52)	
≤26	12 (23.08)
27-30	15 (28.85)
31-33	8 (15.38)
34-36	15 (28.85)
≥37	2 (3.85)
Weight, kg (n=43)	
Median (IQR)	77 (26)
Body mass index, kg/m <sup>2</sup> (n=43)	
Median (IQR)	28.36 (9.88)
Preeclampsia, n (%)	1 (1.96%)
Hypothyroidism, n (%)	18 (35.29%)
Hypertension, n (%)	5 (10.00%)
Diabetes mellitus, n (%)	9 (17.65%)
Severity, n (%)	
1 - Mild ilness	9 (17.31%)
2 - Moderate illness	25 (48.08%)
3 - Severe illness	17 (32.69%)
4 - Critical illness	1 (1.92%)
Length of hospitalisation, days (n=52)	
Median (IQR)	8 (6)
Percentage of lung involvement on CT, % (n=34)	20 (11)
Median (IQR)	20 (11)
Time from the onset of COVID-19 symptoms, days (n=51) Median (IQR)	7 (6)
Oxygen flow, l/min (n=52)	
Median (IQR)	4 (4.5)
Need for oxygen supplementation, n (%)	42 (80.77%)
Need for invasive ventilation, n (%)	0 (0.00%)
Need for high-flow oxygen therapy, n (%)	9 (17.31%)
Need for ICU admission, n (%)	2 (3.85%)
Hemoglobin (g/dl) (n=52)	
Median (IQR)	12 (1.1)
Leukocytes, $\times 10^3/\mu l$ (n=52)	
Median (IQR)	7.7 (3.23)
Lymphocytes, ×10 <sup>3</sup> /µl (n=52)	
Median (IQR)	0.97 (0.32)
Neutrophils, $\times 10^3/\mu l$ (n=52)	
Median (IQR)	5.8 (3.11)
Platelets $(\times 10^3/\mu l)$	
Median (IQR)	184.5 (58.5)
ALP, U/L (n=42)	
Median (IQR)	117 (63)
AST, U/l (n=52)	
Median (IQR)	28 (18.5)
APTT, s (n=49)	
Median (IQR)	33.7 (5.5)
ALT, U/l (n=50)	
Median (IQR)	24 (24)
Bilirubin, mg/dl (n=49)	
Median (IQR)	0.46 (0.51)
NT pro-BNP, pg/ml (n=44)	
Median (IQR)	29.5 (39)

(Contd...)

# TABLE 1. (Continued)

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Variable	Cases	n flo	(in)	ue**	c.	20		01		2	Ľ		33	
Calcium, mmol/l (n=43)		ygeı	(l/m	-val	i	0./		<0.0>		0	5		0.0	
Mean±SD	2.16±0.09	ŏ		d										
Total cholesterol, mg/dl			uo											
Mean±SD	188.51±56.09	h of	rs) rs)	Je**	9	77		ŝ		07	2		3	
CK, U/l (n=50)		engt	itali (day	-valı	Ì	0.7		0.8		0 00	5		0.37	
Median (IQR)	61.5 (69)	Ĕ	dsou	<i>b</i>										
CRP, mg/l (n=52)			2											
Median (IQR)	46.95 (52.4)	Е	of 19 days	44										
D-dimer, µg/l FEU (n=32)		e fro	nset 'ID- ms (	ılue	ļ	Ę		24		-	-		502	
Median (IQR)	1178 (515.5)	Līme	DOV DV ptoi	p-va	(	Ö		0					0.0	
Ferritin, $ng/ml$ (n=50)			tl C sym											
Median (IQR)	83.5 (109)			*.										
Fibrinogen, mg/dl (n=50)	552 (225)			alue	c I	744		936		173	۲ F		011	
Median (IQR)	553 (225)			V-d	¢	0.		0		0	5		0	
GGIP, U/L (n=50)	1((20))				<			()					(9	
Median (IQR)	16 (20)	(1-4		4	000	.03% (%)		22%	(%)	(706,	(%) (%)	5	38%	(%)
Glucose, mg/dl (n=41)	02 (17)	-19			( ,	0(0		1	0	-	- 0	>	1 (2	0
$\frac{1}{2} \frac{1}{2} \frac{1}$	92 (17)	QD			<	<u> </u>		(%			_		()	
Creatinine, mg/di (n=50)	0.52 (0.17)	lõ.			ć	0.3% .33%		89%	0%)	(%)(2)	(%)U	222	.43%	.78%
Lastata debudrogenasa LU/L (n. 50)	0.52 (0.17)	of (			0	0 (3 (33		3 (28	2 (4	5) 91	1 (5	) A	(21)	12
Modian (IOP)	226 5 (01)	erity				- 9		61					6	
Magnesium $mg/dl(n=46)$	220.3 (91)	seve			200	4%)		1%	9	(70	à c		4%)	1%)
Median (IOR)	174(016)	urse		7	1 , 1	51.5 4.4		51.1	(40)	(EO	200		57.1	
Lactates $\text{mmol}/((n-42))$	1.74 (0.10)	Co			Ĩ	9 17 (5 8 (4		23 (	2	ц С	) C	,	24 (	=
Median (IOR)	1 17 (0 71)				ŝ			9					<u></u>	<u></u>
Procalcitonin $ng/ml(n=51)$	1.17 (0.71)			_	1	.15% .22%		.78%	S0%)	(709	(%)) 20%)	2 2	059	.11
Median (IOR)	0 14 (0 19)					6 (15 F (22		3(17	10	0	 	-	3 (15	Ξ
INR (n=51)	0.11(0.17)					11 7		~						
Median (IOR)	1 (0.08)			lue*	c.	8		8		2	,		21	
Total protein, g/dl (n=51)	(0.00)	sion		0-Va	Ċ	0		0.0		C	3		0.2	
Median (IOR)	6.13 (0.6)	mis			,	_								
Urea, mg/dl (n=52)		Jad		es	0	90 (%)		44%	0%)	(707)	1%) 1%	20	2%)	1%)
Median (IQR)	11 (5)	[]		X		(0) (0)		2 (4.	0	c C	- U - U	>	1.0	1
Uric acid, mg/dl (n=50)		for			<			(1 (1)						
Median (IQR)	4.2 (1.7)	leed		0	-			.56%	0%)	(709	0%)	222	8%)	(%6
Vitamin D3 25(OH), ng/ml (n=45)				Z	00)	8 (Y2		(95	[]	0/ 8	200	, , ,	1 (9	8 (8)
Mean±SD	27.15±10.82				č	1 2		4	1(1)			1	7	
Interleukin 6, pg/ml (n=52)		on		le*										
Median (IQR)	26.5 (31.6)	ilati		valt	,	-		-		-	-		1	:
High-sensitive troponin I, ng/ml (n=52)		vent		p-d										
Median (IQR)	2.75 (2.15)	eve		SS	0.00	(%)		(%(	(%(	(70(	(%)	í el	(%(	(%)
ICU: Intensive care unit; BMI: Body mass inde	ex; ALP: Alkaline phos-	9 S(		X	0			0 ((	))	0 ((	20	>	0 ((	0
phatase; AST: Aspartate aminotransferase; A	PPT: Activated partial	for i			1	8		(%(	8	(70(	101	Ś	(%(	(%
thromboplastin time; ALT: Alanine aminotrans	sferase; CK: Creatinine	Sed 1		No	000	(10C		(100	100	0100		2	(100	100
kinase; CRP: C-reactive protein; GGTP: Gamm	a-glutamyl transpepti-	วี   zื			0	18		45	.0	202	3 0	ĩ	42	6
ratio: IL-6: Interleukin 6: Hs-Tnl: High-sensitivi	ity troponin l	py		്പ		_				_			_	
		es a		valu	0	.53(		0.9		500	ŝ		.020	
CRP ferritin II-6 glucose IDH proc	alcitonin (PCT) and	an th		p-v	(	0				0	2		0	
		xyge			ŝ	() ()		(%	_	_		_	_	
high-sensitivity (hs) troponin I predict	ed a severe course of			Yes	ì	[5.5] 2.22		7.78	20%	180/	~~~ (0%)	( ) )	12%	44%
illness as measured by disease course c	lassification, the need	r cc		Ĺ	1	5 (J 4 (2		8 (1	-	0	$\langle \circ \rangle$	>	5	4
for oxygen supplementation, higher der	mand for oxygen sup-	n o' high			_	<u></u>		(%		_				
plementation length of hospital ctay th	he need for mechani-	l for		9	=51	4.85 7.78 9	((	2.22	30%)	3700	.%UC	2	88%	(%9)
plenetilation, length of hospital stay, u		Jeec			n (n	8 (8 4 (7,	n=5(	7 (8:	4 (8	9 FF	5-1 0 (1(	· · _	37 (	5 (£
cal ventilation, and ICU admission. The	e results are presented	Ď]∠o			idisr	1 7	ı) uc	ŝ	í	52) ,		=51)		;
in Table 3. Patients' characteristics an	d laboratory markers	N	e		iyro.		ensi		``	a (n=		ss (n		•
compared across four severity catego	ries are presented in		iabl		potŀ	es es	pert	Jo	es -	un d		bete	lo I	és
Table 4. Univariate logistic regression i	revealed that diabetes	IA	Vau		Η	L Y	Hy	4	~ ·	AS	. >	Dia	4	$\left  \right $

Pearson's Chi-square test, \*\*U-Mann-Whitney test-corrected for continuity

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Variable	Need for oxygen supplementation			Need for high-flow oxygen therapy			Need for ICU admission			Length of hospitalization (days)			Oxygen flow (l/min)			
	Yes	No	p-value*	Yes	No	<i>p</i> -value*	Yes	No	p-value*	n	р	p-value**	n	р	p-value**	
Weight	35	8	0.302	36	7	0.236	42	1	< 0.001	43	0.123	0.433	43	0.252	0.103	
Body mass index	35	8	0.302	36	7	0.104	42	1	< 0.001	43	0.131	0.403	43	0.283	0.066	
Age	42	10	0.918	43	9	0.475	50	2	0.73	52	0.026	0.856	52	-0.135	0.341	
Length of hospitalization (days)	42	10	< 0.001	43	9	< 0.001	50	2	0.183				52	0.65671	< 0.001	
Percentage of lung involvement on CT (%)	31	3	0.237	27	7	0.013	33	1	< 0.001	34	0.321	0.064	34	0.445	0.008	
Hemoglobin	42	10	0.591	43	9	0.195	50	2	0.386	52	-0.253	0.07	52	-0.163	0.248	
Leukocytes	42	10	0.758	43	9	0.52	50	2	0.41	52	-0.252	0.0717	52	-0.055	0.7	
Lymphocytes	42	10	0.021	43	9	0.058	50	2	0.573	52	-0.3	0.031	52	-0.471	< 0.001	
Neutrophils	42	10	0.623	43	9	0.924	50	2	0.317	52	-0.144	0.309	52	0.077	0.585	
Platelets	42	10	0.265	43	9	0.431	50	2	0.045	52	-0.2	0.164	52	-0.14	0.323	
ALP	34	8	0.912	35	7	0.947	40	2	0.794	42	-0.082	0.604	42	-0.123	0.438	
AST	42	10	0.706	43	9	0.46	50	2	0.905	52	0.415	0.002	52	0.153	0.28	
APTT	39	10	0.064	40	9	0.97	47	2	0.034	49	0.109	0.456	49	0.2	0.168	
ALT	40	10	0.409	41	9	0.318	48	2	0.864	50	0.233	0.103	50	0.075	0.607	
Bilirubin	39	10	0.171	40	9	0.352	47	2	0.463	49	0.042	0.777	49	-0.006	0.965	
NT pro-BNP	35	9	0.819	38	6	0.514	43	1	< 0.001	44	0.074	0.633	44	0.029	0.851	
Calcium	34	9	0.22	36	7	0.176	41	2	0.199	43	-0.421	0.005	43	-0.491	< 0.001	
Total cholesterol	32	9	0.004	35	6	0.268	40	1	< 0.001	41	-0.401	0.009	41	-0.543	< 0.001	
СК	40	10	0.641	41	9	0.901	48	2	0.32	50	0.065	0.653	50	0.097	0.502	
CRP	42	10	< 0.001	43	9	0.111	50	2	0.633	52	0.28	0.044	52	0.511	< 0.001	
D-dimer	31	1	< 0.001	23	9	0.869	30	2	0.847	32	-0.154	0.399	32	-0.101	0.583	
Ferritin	40	10	0.056	41	9	0.054	48	2	0.216	50	0.423	0.002	50	0.328	0.02	
Fibrinogen	40	10	0.641	41	9	0.71	48	2	0.392	50	-0.249	0.082	50	-0.041	0.778	
Gamma-glutamyl	40	10	0.074	41	9	0.44	48	2	0.826	50	0.206	0.151	50	0.168	0.242	
Chusese	22	0	0.012	26	~	0.002	41	0	.0.001	41	0.200	0.012	41	0510	0.001	
Giucose	3Z 40	9	0.012	30	5	0.005	41	0	< 0.001	41	0.388	0.012	41	0.518	<0.001	
Creatinine	40	10	0.765	41	9	0.502	48	2	0.2/6	50	-0.192	0.182	50	-0.136	0.347	
Lactate denydrogenase	40	10	0.022	41	9	0.0244	48	2	0.367	50	0.426	0.002	50	0.374	0.007	
Magnesium	37	9	0.311	37	9	0.683	44	2	0.816	40	0.187	0.214	40	0.127	0.402	
Lactates	33	9	0.092	37	5	0.52	42	0	< 0.001	42	0.276	0.077	42	0.283	0.07	
Procaicitonin	41	10	0.001	42	9	0.005	49	2	0.056	51	0.509	< 0.001	51	0.573	<0.001	
	41	10	0.419	42	9	0.118	49	2	0.48	51	0.1	0.495	51	0.305	0.029	
lotal protein	41	10	0.019	42	9	0.069	49	2	0.245	51	-0.406	0.003	51	-0.485	<0.001	
Urea	42	10	0.149	43	9	0.417	50	2	0.018	52	0.046	0.744	52	-0.114	0.42	
Uric acid	40	10	0./11	42	8	0.631	49	1	< 0.001	50	-0.036	0.804	50	-0.129	0.372	
Vitamin D3 25(OH)	35	10	0.697	39	6	0.987	45	0	< 0.001	45	0.035	0.818	45	0.052	0.734	
Iroponin I	42	10	0.365	43	9	0.156	50	2	0.386	52	0.111	0.435	52	0.285	0.041	
Interleukin 6	42	10	<0.001	43	9	0.018	50	2	1	52	0.422	0.002	52	0.618	<0.001	
Time from the onset of COVID-19 symptoms (days)	41	10	0.055	43	8	0.55	49	2	0.98	51	0.257	0.069	51	0.218	0.124	
Oxygen flow (l/min)	42	10	< 0.001	43	9	< 0.001	50	2	0.030	52	0.658	< 0.001				
High-flow oxygen therapy – flow (l/min)	9	0	< 0.001	0	9	< 0.001	7	2	0.889	9	0.008	0.983	9			
High-flow oxygen therapy – FiO	9	0	< 0.001	0	9	< 0.001	7	2	0.667	9	-0.068	0.862	9			

TABLE 3. Correlations of patient's general and clinical characteristics and COVID-19 outcomes

\*U-Mann–Whitney test corrected for continuity; \*\*Spearman test. ICU: Intensive care unit; BMI: Body mass index; ALP: Alkaline phosphatase; AST: Aspartate aminotransferase; APPT: Activated partial thromboplastin time; ALT: Alanine aminotransferase; CK: Creatinine kinase; CRP: C-reactive protein; GGTP: Gamma-glutamyl transpeptidase; LDH: Lactate dehydrogenase; PT: Prothrombin time; IL-6: Interleukin 6

(odds ratio [OR] 10.18, 95% CI 1.83-56.54; p = 0.008), gestational age < 32 weeks (OR 5, 95% CI 1.36-18.43; p = 0.016), lung involvement on CT imaging > 20% (OR 5.8, 95% CI 1.54-21.81, p = 0.009), lymphocyte count < 1(x10<sup>3</sup>/µl) (OR 27.43, 95% CI 3.26-231.58; p = 0.002), calcium level ≤ 2.15 (mmol/l) (OR

5.56, 95% CI 1.61-19.22; p = 0.007), CRP > 75(mg/l) (OR 9.11, 95% CI 2.38-34.85; p = 0.001), IL-6 > 60 (pg/ml) (OR 16.5, 95% CI 1.8-151.58; p = 0.013), procalcitonin > 0.2(ng/ml) (OR 5.11, 95% CI 1.49-17.56; p = 0.010), LDH > 270 (U/l) (OR 3.73, 95% CI 1.04-13.45; p = 0.044), total cholesterol  $\leq$  180 (mg/dl) (OR

ABLE 4. Association of patients	' characteristics and COVID-19 severity
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M	Course severity of COVID-19 (1-4)									
Variable	1	2	3-4	Н	<i>p</i> -value					
Age, years	32 (31-35)	33 (29-33)	31 (29-35)	0.406	0.816					
Gestational age, weeks	33 (26-35)	32 (29-34)	28 (26-31)	4.433	0.109					
Weight, kg	72.5 (61.5-86.5)	76 (71-84)	92.5 (68-102)	3.099	0.212					
Body mass index, kg/m <sup>2</sup>	25.72 (22.72-31.72)	27.43 (25.65-30.08)	33.71 (26.17-37.64)	3.69	0.158					
Percentage of lung involvement on CT, %	-	16.5 (10-20)	25 (19.5-41)	0	$1 (p^{2/3-4}=0.028)$					
Hemoglobin, g/dl	12 (11.9-12.4)	12 (11.6-12.6)	11.65 (11-12.1)	2.727	0.256					
Leukocytes, ×10³/µL	7.7 (5.4-8.95)	7.75 (6.59-9.34)	7.73 (5.21-8.88)	0.586	0.746					
Lymphocytes, $\times 10^3/\mu$ L	1.12 (0.88-1.32)	1.1 (0.92-1.1)	0.85 (0.8-0.93)	10.702	$0.005 (p^{2/3-4}=0.006)$					
Neutrophils, ×10 <sup>3</sup> / $\mu$ L	5.53 (4.19-6.97)	5.78 (5.1-7.81)	6.09 (3.98-7.5)	0.883	0.643					
Platelets, $\times 10^3/\mu L$	188 (179-192)	191 (165-249)	176.5 (145-200)	1.324	0.515					
Alkaline phosphatase, U/l	112 (95-165)	137 (95-158)	94 (89-126)	3.145	0.207					
Aspartate aminotransferase, U/l	26 (19-28)	31 (22-39)	26 (19-54)	1.908	0.385					
Activated partial thromboplastin time, s	31.5 (31.2-35.6)	34.1 (31.7-35.8)	34.35 (32-39.45)	1.352	0.507					
Alanine aminotransferase, U/l	30 (19-31)	22 (12.5-40.5)	26 (13-59)	0.236	0.889					
Bilirubin, mg/dl	0.55 (0.21-0.81)	0.38 (0.3-0.78)	0.53 (0.26-0.71)	0.017	0.991					
NT pro-BNP, pg/ml	31 (23-39.5)	29 (16-48)	44 (16-128)	0.277	0.871					
Calcium, mmol/l	2.22 (2.16-2.29)	2.19 (2.12-2.22)	2.11 (2.07-2.14)	9.875	$0.007 (p^{1/3-4}=0.020; p^{2/3-4}=0.027)$					
Total cholesterol, mgdl	221 (203-262.5)	196 (180-206)	160.5 (122-194)	11.051	$0.004 (p^{1/3-4}=0.003)$					
Creatine kinase, U/l	62 (21-78)	60 (27-95.5)	56 (32-99)	0.061	0.970					
C-reactive protein, mg/l	18.4 (11.4-40.8)	48 (38.3-66.3)	86.15 (36.9-126.5)	12.980	$0.002 \ (p^{1/2}=0.045; p^{1/3.4}<0.001)$					
D-dimer, µg/l FEU	1721 (1046-2396)	1186.5 (1029-1396)	1088.5 (828-1386)	1.288	0.525					
Ferritin, ng/ml	59 (37-72)	84 (50.5-146.5)	130 (75-188)	6.761	$0.034 (p^{1/3-4}=0.028)$					
Fibrinogen, mg/dl	410 (392-596)	590.5 (457.5-663)	519 (467-683)	2.41	0.299					
Gamma-glutamyl transpeptidase, U/l	10 (6-16)	19 (11.5-29.5)	17 (9-33)	3.223	0.200					
Glucose, mg/dl	84 (80-92)	90 (86-97)	101.5 (94-113)	7.627	$0.022 (p^{1/3-4}=0.027)$					
Creatinine, mg/dl	0.51 (0.49-0.65)	0.52 (0.47-0.61)	0.5 (0.41-0.6)	1.174	0.556					
lactate dehydrogenase, U/l	203 (189-210)	235 (195.5-267.5)	245 (198-289)	4.039	0.132					
Magnesium, mg/dl	1.69 (1.67-1.76)	1.74 (1.64-1.8)	1.77 (1.67-1.98)	1.451	0.484					
Lactates, mmol/l	1 (0.82-1.31)	1.17 (0.95-1.68)	1.33 (0.95-1.86)	2.183	0.336					
Procalcitonin, ng/ml	0.07 (0.05-0.08)	0.14 (0.09-0.22)	0.24 (0.14-0.5)	16.626	<0.001 ( $p^{1/2}$ =0.026; $p^{1/3-4}$ <0.001)					
Prothrombin time, s	1.01 (0.96-1.04)	0.98 (0.97-1.01)	1.05 (1-1.11)	7.013	$0.030 (p^{2/3-4}=0.025)$					
Total protein, g/dl	6.43 (6.13-6.66)	6.34 (6.04-6.47)	5.84 (5.65-6.1)	13.283	$0.001 \ (p^{1/3.4} = 0.011; p^{1/34} = 0.003)$					
Urea, mg/dl	12 (11-14)	11 (9-14)	11 (8-14)	1.040	0.595					
Uric acid, mg/dl	4.3 (3-4.6)	4.2 (3.6-5.7)	3.8 (2.7-4.75)	3.696	0.158					
Vitamin D3 25(OH), ng/ml	25.2 (23-33.6)	29.1 (16-34.6)	26 (19.1-30)	0.907	0.635					
Troponin I, ng/ml	1.4 (1.1-2)	2.8 (0.8-3.2)	3.2 (1.6-6.8)	6.624	0.036					
Interleukin 6, pg/ml	15.2 (5.92-17)	28 (14.6-36.9)	46.55 (22.9-65.7)	12.509	$0.002 \ (p^{1/3-4}=0.001)$					

Data are presented as median (interquartile ranges). *P* values were calculated using the Kruskal–Wallis test (df=2 for all variables) followed by Dunn's *post hoc* test.  $p^{1/2}$  values indicate the comparison between course severity 1 (mild disease) and 2 (moderate disease);  $p^{1/34}$  values indicate the comparison between course severity 1 (mild disease) and 3-4 (severe or critical disease);  $p^{2/34}$  values indicate the comparison between course severity 2 (moderate disease) and 3-4 (severe or critical disease). ICU: Intensive care unit; BMI: Body mass index; ALP: Alkaline phosphatase; AST: Aspartate aminotransferase; APPT: Activated partial thromboplastin time; ALT: Alanine aminotransferase; CK: Creatinine kinase; CRP: C-reactive protein; GGTP: Gamma-glutamyl transpeptidase; LDH: Lactate dehydrogenase; PT: Prothrombin time; IL-6: Interleukin 6

5.65, 95% CI 1.53-20.93; p = 0.010), total protein level  $\leq$  6.3 (g/dl) (OR 9, 95% CI 1.79-45.34; p = 0.008), hs-troponin I > 6 (ng/ml) (OR 12.69, 95% CI 1.35-119.34), and glucose > 99 (mg/dl) (OR 6, 95% CI 1.48-24.27; p = 0.012) were associated with increased risk of severe-to-critical COVID-19.

## DISCUSSION

In our study, the median date of the pregnant women's hospital admission was the  $30^{th}$  gestational week (range =  $17-37^{th}$  week). Because COVID-19 is an immune-mediated intracellular viral infection, it may pose a threat during pregnancy due to the special immunological adaptations that improve a pregnant woman's tolerance to the fetal semi-allograft late in the second trimester and the increased inflammatory response in the third trimester [10,14,23-25]. In addition, hypertension, diabetes, and cardiovascular diseases that develop during the third trimester may predispose pregnant women to the severe course of illness. Therefore, we advise vaccination in the second trimester for maternal and fetal benefits [26].

We found that median lung involvement was 20%, with a range of 1-60\%, and as lung involvement at the time of admission correlated with 4 of the 6 main outcomes – the severity

of the course of the COVID-19 disease, oxygen flow (l/min), the need for high-flow oxygen therapy, and the need for ICU admission – lung involvement may be considered as a predictor of disease aggravation. Based on the pathophysiology of COVID-19 progression, it seems like a truism to claim that the greater lung involvement is, the greater the severity of the disease, which is corroborated in our findings. However, the previous studies in pregnant women report contrasting results: One indicating greater lung involvement among pregnant women, and another reporting that lung involvement was similar in both pregnant and non-pregnant subjects [27,28].

Several previous studies have attempted to determine which laboratory parameters correlate with disease severity among pregnant women with COVID-19. Those studies found that subjects' laboratory results largely mirrored those in the adult non-pregnant population, especially regarding lymphopenia and inflammation parameters.

Severe COVID-19 is associated with higher levels of inflammatory markers than in mild disease. Therefore, tracking these markers may permit early identification of patients at risk of disease progression. Likewise, a link between increased cardiac markers and disease aggravation with a few potential pathomechanisms is well established in the literature [29]. COVID-19 can cause direct or indirect heart injury: cardiomyocyte viral infection, cytokine-mediated systemic inflammation, supply-demand mismatch, and micro- and macrovascular thrombosis [29-31].

Lymphopenia has been identified as the most distinctive predictive parameter [32-34]. In a study by Lombardi et al., lymphocyte values at admission correlated with the oxygen need. CRP levels were found to be the inflammatory biomarker that better mirrored the course of the disease than D-dimer or ferritin levels, which were not reliable predictors of a poor outcome [32]. The retrospective study of 217 pregnant women with COVID-19 by Bozkurt et al. showed that elevated LDH, CRP, IL-6, and ferritin levels coupled with low albumin levels on hospital admission were predictive parameters for a more severe course of illness, and that elevated serum levels of blood urea nitrogen and creatine were the most predictive parameters for ICU admission [35]. Data support that the host's immune system overreaction (cytokine storm syndrome or cytokine release syndrome) may play an important role in the pathogenesis of SARS [36]. SARS-CoV infection may lead to hyper-induction of the immune system, causing increased levels of cytokines, e.g., IL-6 and chemokines, all of which have been observed in SARS patients. However, there are also contradicting results [37]. Data have shown that IL-6 levels are also significantly higher in COVID-19 patients with severe disease compared with those with a

non-severe condition. Therefore, IL-6 is a prognostic marker for serious COVID-19 cases in pregnant [38] and non-pregnant cohorts [39-41].

Our study identified a positive correlation between exact glucose values at admission and poorer patient outcomes. This observation suggests that the elevated blood sugar levels we observed may be the result of physiological stress triggered by the disease. COVID-19 disrupts glucose regulation, rendering poor glycemic control, and necessitating particularly careful management in patients with diabetes [42,43]. Indeed, prior work has shown that even in cases of well-controlled pre-existing diabetes, hyperglycemia was commonly observed in acutely ill hospitalized patients and linked to adverse outcomes [44,45]. It seems that COVID-19 may lead to high blood glucose levels in patients with normal glycemic status by modulating immune and inflammatory responses, directly affecting morbidity and mortality [46-48]. In a study by Charoenngam et al. in patients without a history of diabetes, hyperglycemia on the day of admission was shown to have a statistically significant association with mortality, ICU admission, intubation, acute kidney injury, and severe sepsis/septic shock, after adjusting for potential confounders. Therefore, it could be a strong indicator of a high inflammatory burden, leading to a higher risk of severe COVID-19 [49]. Thereby, we recommend that clinicians pay more attention to the blood glucose status of pregnant women with COVID-19, even those who may not have been diagnosed with diabetes prior to admission.

In our study, calcium serum levels were negatively correlated with three measured clinical outcomes: the length of hospitalization (days), the severity of the course of COVID-19 (1-4), and oxygen flow (l/min). These findings were consistent with previously published reports which have showed that low serum calcium levels are associated with disease severity and a poor prognosis for patients with COVID-19 [50-53]. In a study by Zhang et al. low serum calcium levels were the most predictive feature of COVID-19 diagnosis of all models tested [54]. The cause of hypocalcemia in COVID-19 patients is not clear. It is commonly found in the laboratory results in patients diagnosed with viral infections and pneumonia [55], and several mechanisms may be suggested. Firstly, the pro-inflammatory cytokines in COVID-19 patients inhibit parathyroid hormone (PTH) secretion, and the resulting impaired response to PTH causes an imbalance of calcium levels [56]. According to the previous studies, levels of the disease progression indicators CRP, PCT, IL-6, and D-dimer are found to be significantly higher in COVID-19 patients with hypocalcemia. When this is coupled with calcium serum levels which are negatively correlated with these indicators, it means that these patients may have

a greater inflammatory response [50,51,53]. Secondly, the occurrence of hypocalcemia may be associated with calcium inflow due to hypoxemic tissue damage. Another theory is that modification of calcium levels is crucial for the survival and replication of the SARS-CoV-2, since calcium is used in virus structure formation, entry, gene expression, virion maturation, and release [57]. Pregnancy also often leads to vitamin D deficiency resulting in hypocalcemia due to impaired intestinal absorption and thus an inadequate intake of calcium [58]. Finally, calcium is predominantly bound to albumin in plasma, and a decrease in serum albumin or total protein levels, mainly occurring in the third trimester, will cause hypocalcemia [59].

Our work showed that low total protein serum levels are a predictive factor for both a longer time to clinical improvement and a greater severity of disease, and as such this predictor can provide useful information for clinicians caring for pregnant women with COVID-19. Several mechanisms were proposed, including anti-oxidative and anti-inflammatory values of albumin [60,61], downregulation of albumin and prealbumin caused by the cytokine storm [62], and dysregulation of the immune system triggered by low protein serum levels [63]. The previous studies have indicated that serum albumin [64-66], prealbumin [62], and total protein levels are poor prognosis parameters among non-pregnant cohorts [63]. As mentioned earlier, total protein and albumin levels decrease because of the physiological processes of pregnancy during the third trimester. This condition poses a major threat to pregnant SARS-CoV-2 infected women as outlined in a study by Bozkurt et al. [35].

Our study showed that on admission, disrupted total cholesterol levels correlate with a greater severity of disease and with five out of the six main outcomes: the length of hospitalization (days), the severity of the course of COVID-19 (1-4), oxygen flow (l/min), the need for oxygen supplementation, and the need for ICU admission. The role of cholesterol in immunity is well established in numerous observational studies. In addition, dynamic changes in lipid levels caused by SARS-CoV-2 might be explained by several hypotheses. Firstly, the production of apolipoproteins and lipoproteins might be impaired by liver damage [67] and cytokine activity [68], and secondly, capillary leakage may occur, relocating them to extravascular compartments [69]. A study by Wei et al. demonstrated that patients with COVID-19 develop hypolipidemia during early stages of the disease and that abnormalities in lipid metabolism progressively became worse in association with the severity of the disease [70]. Lower levels of total cholesterol, low-density lipoprotein, and high-density lipoprotein (HDL) were linked to higher mortality rates and poorer prognoses in patients with COVID-19 [70-73]. Moreover, high CRP/HDL-C ratios were established as an independent

predictor of in-hospital mortality [74]. However, none of these previously published studies involved pregnant women.

The main limitations of our investigation include its single-center nature, as well as its small and homogeneous cohort of patients. Moreover, we evaluated markers on admission, and not their response to disease progression and treatment.

## CONCLUSION

Pregnant women with COVID-19 were hospitalized during their second or third trimester, with dyspnea, cough, and fever as leading symptoms. Concomitant conditions, including diabetes and hypertension, modified the course of illness. CT chest scan at initial presentation may enable medical services required by pregnant patients infected with SARS-CoV-2 to be prioritized. Lymphocytopenia, hypocalcemia, low total cholesterol, low total protein levels, and high serum levels of CRP, ferritin, IL-6, procalcitonin, hs-troponin I, LDH, and glucose measured on hospital admission are good predictors of disease severity and may lead to early identification of patients at risk for developing complications, thereby improving optimization and prevention efforts in this cohort. Further, research with a larger patient sample and risk models is needed to provide useful information for effective health resource management during the COVID-19 pandemic.

## REFERENCES

- WHO Coronavirus (COVID-19) Dashboard. Available from: https://www.covid19.who.int [Last accessed on 2022 Apr 18].
- [2] Breslin N, Baptiste C, Gyamfi-Bannerman C, Miller R, Martinez R, Bernstein K, et al. Coronavirus disease 2019 infection among asymptomatic and symptomatic pregnant women: Two weeks of confirmed presentations to an affiliated pair of New York City hospitals. Am J Obstet Gynecol MFM 2020;2(2):100118.
  - http://doi.org/10.1016/j.ajogmf.2020.100118
- [3] Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet 2020;395(10223):497-506. http://doi.org/10.1016/S0140-6736(20)30183-5
- [4] Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, et al. China medical treatment expert group for covid-19. Clinical characteristics of coronavirus disease 2019 in China. N Engl J Med 2020;382(18):1708-20. http://doi.org/10.1056/NEJM0a2002032
- [5] Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. JAMA 2020;323(11):1061-9. http://doi.org/10.1001/jama.2020.1585
- [6] Grant MC, Geoghegan L, Arbyn M, Mohammed Z, McGuinness L, Clarke EL, et al. The prevalence of symptoms in 24,410 adults infected by the novel coronavirus (SARS-CoV-2; COVID-19): A systematic review and meta-analysis of 148 studies from 9 countries. PLoS One 2020;15(6):e0234765. http://doi.org/10.1371/journal.pone.0234765
- [7] Rogers JP, Chesney E, Oliver D, Pollak TA, McGuire P, Fusar-Poli P,

et al. Psychiatric and neuropsychiatric presentations associated with severe coronavirus infections: A systematic review and meta-analysis with comparison to the COVID-19 pandemic. Lancet Psychiatry 2020;7(7):611-27.

http://doi.org/10.1016/S2215-0366(20)30203-0

- [8] Nowacka U, Kozlowski S, Januszewski M, Sierdzinski J, Jakimiuk A, Issat T. COVID-19 pandemic-related anxiety in pregnant women. Int J Environ Res Public Health 2021;18(14):7221. http://doi.org/10.3390/ijerph18147221
- [9] Kotsev SV, Miteva D, Krayselska S, Shopova M, Pishmisheva-Peleva M, Stanilova SA, et al. Hypotheses and facts for genetic factors related to severe COVID-19. World J Virol 2021;10(4):137-55. http://doi.org/10.5501/wjv.v10.i4.137
- [10] Ferrer-Oliveras R, Mendoza M, Capote S, Pratcorona L, Esteve-Valverde E, Cabero-Roura L, et al. Immunological and physiopathological approach of COVID-19 in pregnancy. Arch Gynecol Obstet 2021;304(1):39-57.

http://doi.org/10.1007/s00404-021-06061-3

- [11] Chen H, Guo J, Wang C, Luo F, Yu X, Zhang W, et al. Clinical characteristics and intrauterine vertical transmission potential of COVID-19 infection in nine pregnant women: A retrospective review of medical records. Lancet 2020;395(10226):809-15. http://doi.org/10.1016/S0140-6736(20)30360-3
- [12] Liu H, Wang LL, Zhao SJ, Kwak-Kim J, Mor G, Liao AH. Why are pregnant women susceptible to COVID-19? An immunological viewpoint. J Reprod Immunol 2020;139:103122. http://doi.org/10.1016/j.jri.2020.103122
- [13] Rasmussen SA, Jamieson DJ, Uyeki TM. Effects of influenza on pregnant women and infants. Am J Obstet Gynecol 2012;207(3):S3-8. http://doi.org/10.1016/j.ajog.2012.06.068
- [14] Mor G, Aldo P, Alvero AB. The unique immunological and microbial aspects of pregnancy. Nat Rev Immunol 2017;17(8):469-82. http://doi.org/10.1038/nri.2017.64
- [15] Allotey J, Stallings E, Bonet M, Yap M, Chatterjee S, Kew T, et al. for PregCOV-19 living systematic review consortium. Clinical manifestations, risk factors, and maternal and perinatal outcomes of coronavirus disease 2019 in pregnancy: Living systematic review and meta-analysis. BMJ 2020;370:m3320. http://doi.org/10.1136/bmj.m3320
- [16] Ellington S, Strid P, Tong VT, Woodworth K, Galang RR, Zambrano LD, et al. Characteristics of women of reproductive age with laboratory-confirmed SARS-CoV-2 infection by pregnancy status-United States, January 22-June 7, 2020. MMWR Morb Mortal Wkly Rep 2020;69(25):769-75.

http://doi.org/10.15585/mmwr.mm6925a1

[17] Zambrano LD, Ellington S, Strid P, Galang RR, Oduyebo T, Tong VT, et al. CDC COVID-19 response pregnancy and infant linked outcomes team. Update: Characteristics of symptomatic women of reproductive age with laboratory-confirmed SARS-CoV-2 infection by pregnancy status-United States, January 22-October 3, 2020. MMWR Morb Mortal Wkly Rep 2020;69(44):1641-7.

http://doi.org/10.15585/mmwr.mm6944e3

- [18] Schwartz DA, Graham AL. Potential maternal and infant outcomes from (Wuhan) coronavirus 2019-nCoV infecting pregnant women: Lessons from SARS, MERS, and other human coronavirus infections. Viruses 2020;12(2):194. http://doi.org/10.3390/v12020194
- [19] Siston AM, Rasmussen SA, Honein MA, Fry AM, Seib K, Callaghan WM, et al., Pandemic H1N1 Influenza in Pregnancy Working Group. Pandemic 2009 influenza A(H1N1) virus illness among pregnant women in the United States. JAMA 2010;303(15):1517-25.

http://doi.org/10.1001/jama.2010.479

[20] Jamieson DJ, Honein MA, Rasmussen SA, Williams JL, Swerdlow DL, Biggerstaff MS, et al., Novel Influenza A (H1N1) Pregnancy Working Group. H1N1 2009 influenza virus infection during pregnancy in the USA. Lancet 2009;374(9688):451-8.

http://doi.org/10.1016/S0140-6736(09)61304-0

[21] Hause AM, Panagiotakopoulos L, Weintraub ES, Sy LS, Glenn SC. Adverse outcomes in pregnant women hospitalized with respiratory syncytial virus infection: A case series. Clin Infect Dis 2021;72(1):138-40.

https://doi.org/10.1093/cid/ciaa668

[22] Flisiak R, Horban A, Jaroszewicz J, Kozielewicz D, Pawłowska M, Parczewski M, et al. Management of SARS-CoV-2 infection: Recommendations of the Polish association of epidemiologists and infectiologists as of March 31, 2020. Pol Arch Intern Med 2020;130(4):352-7.

https://doi.org/10.20452/pamw.15270

- [23] Figueiro-Filho EA, Yudin M, Farine D. COVID-19 during pregnancy: An overview of maternal characteristics, clinical symptoms, maternal and neonatal outcomes of 10,996 cases described in 15 countries. J Perinat Med 2020;48(9):900-11. https://doi.org/10.1515/jpm-2020-0364
- [24] Littauer EQ, Esser ES, Antao OQ, Vassilieva EV, Compans RW, Skountzou I. H1N1 influenza virus infection results in adverse pregnancy outcomes by disrupting tissue-specific hormonal regulation. PLoS Pathog 2017;13(11):e1006757. https://doi.org/10.1371/journal.ppat.1006757

Kumpel BM, Manoussaka MS. Placental immunology and mater-[25]

- nal alloimmune responses. Vox Sang 2012;102(1):2-12. https://doi.org/10.1111/j.1423-0410.2011.01533.x
- [26] Zdanowski W, Waśniewski T. Evaluation of SARS-CoV-2 spike protein antibody titers in cord blood after COVID-19 vaccination during pregnancy in polish healthcare workers: Preliminary results. Vaccines (Basel) 2021;9(6):675. https://doi.org/10.3390/vaccines9060675
- [27] Abedzadeh-Kalahroudi M, Sehat M, Vahedpour Z, Talebian P, Haghighi A. Clinical and obstetric characteristics of pregnant women with Covid-19: A case series study on 26 patients. Taiwan J Obstet Gynecol 2021;60(3):458-62. https://doi.org/10.1016/j.tjog.2021.03.012
- [28] Liu F, Liu H, Li J, Hou L, Lan W, Wang D. Clinico-Radiological Features and Outcomes in Pregnant Women with COVID-19: Compared with Age-Matched Non-Pregnant Women. Available from: https://www.ssrn.com/abstract=3556647 http://doi.org/10.2139/ssrn.3556647
- [29] Wungu CD, Khaerunnisa S, Putri EA, Hidayati HB, Qurnianingsih E, Lukitasari L, et al. Meta-analysis of cardiac markers for predictive factors on severity and mortality of COVID-19. Int J Infect Dis 2021;105:551-9.

http://doi.org/10.1016/j.ijid.2021.03.008

Sattar Y, Ullah W, Rauf H, Virk HU, Yadav S, Chowdhury M, [30] et al. COVID-19 cardiovascular epidemiology, cellular pathogenesis, clinical manifestations and management. Int J Cardiol Heart Vasc 2020;29:100589.

http://doi.org/10.1016/j.ijcha.2020.100589

[31] Lang JP, Wang X, Moura FA, Siddiqi HK, Morrow DA, Bohula EA. A current review of COVID-19 for the cardiovascular specialist. Am Heart J 2020;226:29-44.

https://doi.org/10.1016/j.ahj.2020.04.025

- [32] Lombardi A, Duiella S, Piani LL, Comelli A, Ceriotti F, Oggioni M, et al. Inflammatory biomarkers in pregnant women with COVID-19: A retrospective cohort study. Sci Rep 2021;11(1):13350. https://doi.org/10.1038/s41598-021-92885-7
- [33] Scott R, Hewitt H, Mallet C, Herd L, Shibley C, Bolger S, et al. Recognition and Treatment of Severe COVID-19 in Pregnancy: Lessons from a Cohort of 69 Infected Women and an Evidencebased Guideline. United States: Authorea; 2020. https://doi.org/10.22541/au.160616173.35255142/v1.
- [34] Andrikopoulou M, Madden N, Wen T, Aubey JJ, Aziz A, Baptiste CD, et al. Symptoms and critical illness among obstetric patients with coronavirus disease 2019 (COVID-19) infection. Obstet Gynecol 2020;136(2):291-9.

https://doi.org/10.1097/AOG.00000000003996.

[35] Bozkurt F, Çoşkun Ö, Yeleç S, Bekçibaşi M, Asena M, Bağli I.

Comparison of the clinical and laboratory findings in COVID-19 positive pregnants without comorbidity. Turk J Med Sci 2021 Jul 8. doi: 10.3906/sag-2105-116. Epub ahead of print https://doi.org/10.3906/sag-2105-116

- [36] Song P, Li W, Xie J, Hou Y, You C. Cytokine storm induced by SARS-CoV-2. Clin Chim Acta 2020;509:280-7. https://doi.org/10.1016/j.cca.2020.06.017
- [37] Zhu Z, Cai T, Fan L, Lou K, Hua X, Huang Z, et al. Clinical value of immune-inflammatory parameters to assess the severity of coronavirus disease 2019. Int J Infect Dis 2020;95:332-9. https://doi.org/10.1016/j.ijid.2020.04.041
- [38] Tanacan A, Yazihan N, Erol SA, Anuk AT, Yucel Yetiskin FD, Biriken D, et al. The impact of COVID-19 infection on the cytokine profile of pregnant women: A prospective case-control study. Cytokine 2021;140:155431.
- https://doi.org/10.1016/j.cyto.2021.155431
- [39] Chen X, Zhao B, Qu Y, Chen Y, Xiong J. Detectable serum SARS-CoV-2 viral load [RNAaemia] is closely correlated with drastically elevated interleukin 6 [IL-6] level in critically ill COVID-19 patients. Clin Infect Dis 2020;71:1937-42.
- [40] Ulhaq ZS, Soraya GV. Interleukin-6 as a potential biomarker of COVID-19 progression. Med Mal Infect 2020;50:382-3. https://doi.org/10.1016/j.medmal.2020.04.002
- [41] Aziz M, Fatima R, Assaly R. Elevated interleukin-6 and severe COVID-19: A meta-analysis. J Med Virol 2020;92(11):2283-5. https://doi.org/10.1002/jmv.25948
- [42] Gianchandani R, Esfandiari NH, Ang L, Iyengar J, Knotts S, Choksi P, et al. Managing hyperglycemia in the COVID-19 inflammatory storm. Diabetes 2020;69(10):2048-53. https://doi.org/10.2337/dbi20-0022
- [43] Feldman EL, Savelieff MG, Hayek SS, Pennathur S, Kretzler M, Pop-Busui R. COVID-19 and diabetes: A collision and collusion of two diseases. Diabetes 2020;69(12):2549-65. https://doi.org/10.2337/dbi20-0032
- [44] Corathers SD, Falciglia M. The role of hyperglycemia in acute illness: Supporting evidence and its limitations. Nutrition 2011;27(3):276-81. https://doi.org/10.1016/j.nut.2010.07.013
- [45] McCowen KC, Malhotra A, Bistrian BR. Stress-induced hyperglycemia. Crit Care Clin 2001;17(1):107-24.
- https://doi.org/10.1016/s0749-0704(05)70154-8
- [46] Miftode E, Miftode L, Coman I, Prepeliuc C, Obreja M, Stămăteanu O, et al. Diabetes mellitus-a risk factor for unfavourable outcome in COVID-19 patients-the experience of an infectious diseases regional hospital. Healthcare (Basel) 2021;9(7):788.

https://doi.org/10.3390/healthcare9070788

- [47] Zhang Y, Li H, Zhang J, Cao Y, Zhao X, Yu N, et al. The clinical characteristics and outcomes of patients with diabetes and secondary hyperglycaemia with coronavirus disease 2019: A single-centre, retrospective, observational study in Wuhan. Diabetes Obes Metab 2020;22(8):1443-54.
- https://doi.org/10.1111/dom.14086
- [48] Wang S, Ma P, Zhang S, Song S, Wang Z, Ma Y, et al. Fasting blood glucose at admission is an independent predictor for 28-day mortality in patients with COVID-19 without previous diagnosis of diabetes: A multi-centre retrospective study. Diabetologia 2020;63(10):2102-11.

https://doi.org/10.1007/s00125-020-05209-1

- [49] Charoenngam N, Alexanian SM, Apovian CM, Holick MF. Association between hyperglycemia at hospital presentation and hospital outcomes in COVID-19 patients with and without type 2 diabetes: A retrospective cohort study of hospitalized inner-city COVID-19 patients. Nutrients 2021;13(7):2199. https://doi.org/10.3390/nu13072199
- [50] Liu J, Han P, Wu J, Gong J, Tian D. Prevalence and predictive value of hypocalcemia in severe COVID-19 patients. J Infect Public Health 2020;13(9):1224-8. https://doi.org/10.1016/j.jiph.2020.05.029
- [51] Sun JK, Zhang WH, Zou L, Liu Y, Li JJ, Kan XH, et al. Serum

calcium as a biomarker of clinical severity and prognosis in patients with coronavirus disease 2019. Aging (Albany NY). 2020;12(12):11287-95.

- https://doi.org/10.18632/aging.103526
- [52] Cappellini F, Brivio R, Casati M, Cavallero A, Contro E, Brambilla P. Low levels of total and ionized calcium in blood of COVID-19 patients. Clin Chem Lab Med 2020;58(9):e171-3. https://doi.org/10.1515/cclm-2020-0611
- [53] Qi X, Kong H, Ding W, Wu C, Ji N, Huang M, et al. Abnormal coagulation function of patients with COVID-19 is significantly related to hypocalcemia and severe inflammation. Front Med (Lausanne) 2021;8:638194.

https://doi.org/10.3389/fmed.2021.638194

- [54] Zhang J, Jun T, Frank J, Nirenberg S, Kovatch P, Huang KL. Prediction of individual COVID-19 diagnosis using baseline demographics and lab data. Sci Rep 2021;11(1):13913. https://doi.org/10.1038/s41598-021-93126-7
- [55] Sankaran RT, Mattana J, Pollack S, Bhat P, Ahuja T, Patel A, et al. Laboratory abnormalities in patients with bacterial pneumonia. Chest 1997;111(3):595-600.

https://doi.org/10.1378/chest.111.3.595

- [56] Fong J, Khan A. Hypocalcemia: Updates in diagnosis and management for primary care. Can Fam Physician 2012;58(2):158-62.
- [57] Wang JM, Liu W, Chen X, McRae MP, McDevitt JT, Fenyö D. Predictive modeling of morbidity and mortality in patients hospitalized with COVID-19 and its clinical implications: Algorithm development and interpretation. J Med Internet Res 2021;23(7):e29514.

https://doi.org/10.2196/29514

- [58] Holick MF. Vitamin D deficiency. N Engl J Med 2007;357(3):266-81. https://doi.org/10.1056/NEJMra070553
- [59] Elango R, Ball RO. Protein and amino acid requirements during pregnancy. Adv Nutr 2016;7(4):839S-44. https://doi.org/10.3945/an.115.011817
- [60] Artigas A, Wernerman J, Arroyo V, Vincent JL, Levy M. Role of albumin in diseases associated with severe systemic inflammation: Pathophysiologic and clinical evidence in sepsis and in decompensated cirrhosis. J Crit Care 2016;33:62-70. https://doi.org/10.1016/j.jcrc.2015.12.019
- [61] Rozga J, Piątek T, Małkowski P. Human albumin: Old, new, and emerging applications. Ann Transplant 2013;18:205-17. https://doi.org/10.12659/AOT.889188
- [62] Akbar MR, Pranata R, Wibowo A, Lim MA, Sihite TA, Martha JW, et al. The association between serum prealbumin and poor outcome in COVID-19-systematic review and meta-analysis. Eur Rev Med Pharmacol Sci 2021;25(10):3879-85. https://doi.org/10.26355/eurrev\_202105\_25955
- [63] Mei Q, Wang AY, Bryant A, Yang Y, Li M, Wang F, et al. Development and validation of prognostic model for predicting mortality of COVID-19 patients in Wuhan, China. Sci Rep 2020;10(1):22451.

https://doi.org/10.1038/s41598-020-78870-6

[64] Acharya R, Poudel D, Patel A, Schultz E, Bourgeois M, Paswan R, et al. Low serum albumin and the risk of hospitalization in COVID-19 infection: A retrospective case-control study. PLoS One 2021;16(4):e0250906.

https://doi.org/10.1371/journal.pone.0250906

- [65] Kheir M, Saleem F, Wang C, Mann A, Chua J. Higher albumin levels on admission predict better prognosis in patients with confirmed COVID-19. PLoS One 2021;16(3):e0248358. https://doi.org/10.1371/journal.pone.0248358
- [66] Xu Y, Yang H, Wang J, Li X, Xue C, Niu C, et al. Serum albumin levels are a predictor of COVID-19 patient prognosis: Evidence from a single cohort in Chongqing, China. Int J Gen Med 2021;14:2785-97.

https://doi.org/10.2147/IJGM.S312521

[67] Poynard T, Deckmyn O, Rudler M, Peta V, Ngo Y, Vautier M, et al. Performance of serum apolipoprotein-A1 as a sentinel of Covid-19. PLoS One 2020;15(11):e0242306. https://doi.org/10.1371/journal.pone.0242306

- [68] Ettinger WH, Varma VK, Sorci-Thomas M, Parks JS, Sigmon RC, Smith TK, et al. Cytokines decrease apolipoprotein accumulation in medium from Hep G2 cells. Arterioscler Thromb 1994;14(1):8-13.
  - https://doi.org/10.1161/01.atv.14.1.8
- [69] van Leeuwen HJ, Heezius EC, Dallinga GM, van Strijp JA, Verhoef J, van Kessel KP. Lipoprotein metabolism in patients with severe sepsis. Crit Care Med 2003;31(5):1359-66. https://doi.org/10.1097/01.CCM.0000059724.08290.51
- [70] Wei X, Zeng W, Su J, Wan H, Yu X, Cao X, et al. Hypolipidemia is associated with the severity of COVID-19. J Clin Lipidol 2020;14(3):297-304. https://doi.org/10.1016/j.jacl.2020.04.008
- [71] Yue J, Xu H, Zhou Y, Liu W, Han X, Mao Q, et al. Dyslipidemia is related to mortality in critical patients with coronavirus disease

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- [72] Zhao M, Luo Z, He H, Shen B, Liang J, Zhang J, et al. Decreased low-density lipoprotein cholesterol level indicates poor prognosis of severe and critical COVID-19 Patients: A retrospective, single-center study. Front Med (Lausanne) 2021;8:585851. https://doi.org/10.3389/fmed.2021.585851
- [73] Lassale C, Hamer M, Hernáez Á, Gale CR, Batty GD. Association of pre-pandemic high-density lipoprotein cholesterol with risk of COVID-19 hospitalisation and death: The UK Biobank cohort study. Prev Med Rep 2021;23:101461. https://doi.org/10.1101/2021.01.20.21250152
- [74] Li Y, Zhang Y, Lu R, Dai M, Shen M, Zhang J, et al. Lipid metabolism changes in patients with severe COVID-19. Clin Chim Acta. 2021;517:66-73.

https://doi.org/10.1016/j.cca.2021.02.011

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